











Institut Pythéas Observatoire des Sciences de l'Univers Aix+Marseille Université

FLOW CYTOMETRY DATA MANAGEMENT

CytoBuoy Meeting 2017

Woerden (Netherlands) – March 27th, 2017

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MIO/CNRS

Cytobuoy FCM MIO deployment



CytoBuoy Meeting – 27- 29 March 2017





FCM tools processing







Barriers to exchange/re-use FCM data

- Functional group names definition
- Metadata missing: Standard Beads, trigger, etc..
- Standard data file schemas and formats
- FCM data quality control (QC)
- FCM standard vocab (34 codes)

FCM instrument and data are missing in EU portals



Barriers to exchange/re-use FCM data



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Data Management?





"Data Resource Management is the development and execution of architectures, policies, practices and procedures that properly manage the full data lifecycle."



MIO FCM Data management (working on interoperability with SeaDataNet)





→ Selected CytoClus output by MIO



🕁 CytoClus3 - 2umR	EDfilteredsw-flr6 2016-10-	-21 06u23.cyz			4				- 0 X
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→ Data Consolidation

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Cytobase Input Processor (Mathilde Dugenne, 2015)

mathilde.dugenne@mio.osupytheas.fr



➔ Example of standardization



SELECTION SET	STANDARDIZED NAMES
Cryptophytes	Cryptophytes
Microphytoplank	Microphytoplankton
largephyto3 - N/mL	Microphytoplankton
My favorite group	Microphytoplankton
largephyto1 - N/mL	Microphytoplankton
Nano1	Nanoeukaryotes
Nanoeukaryote 2s	Nanoeukaryotes

→ Data Consolidation





© Tools developed by M. Dugenne, 2015

→ High resolution flow cytometry data visualisation





FLOW CYTOMETRY DATABASE

Česko. Україна • Paris Slovensko France Moldova Magyarország Switzerland România Hrvatska Србија България Italia España Roma Istanbul Shqipëria Portugal Türkiye Ελλάδα Κύπρος سورية Casablanca الدار البيضاء Dilli الغراق المغرب لأسكندنية • الفاهرة الجزائر مصر معودية Leaflet | Map data @ OpenStreetMap contributors, CC-BY-SA, Imagery @ Mapbox

Pour le trajet de Marseille à Tunis à bord du C/F CARTHAGE Du 2016-03-23 au 2016-03-26

Nombre total des stations :

CytoSense :43



Total Fluorescence Rouge (u.a..cm⁻³)



Total Abondance (cell.cm⁻³)



→ High resolution flow cytometry data visualisation





orange(FLO)'(u.a./cell)

Ecart-type du'Total de la fluorescence

FLOW CYTOMETRY DATABASE



Latitude (°) de Marseille à Tunis



Moyenne du'Total de la fluorescence rouge(FLR)'(u.a./cell)

Toward EU standardisation of FCM data









PI: Patrick FARCY (IFREMER)

PI: Michèle FICHAUT (SISMER/IFREMER)

Task 3.1: Automated platform for the observation of Phytoplankton diversity in relation to ecosystem services



WP9.5.2: Ingesting, validating, long-term storage and access of Flow Cytometer data

Leader: Felipe ARTIGAS (CNRS-ULCO)

Leaders: VLIZ, CNRS-MIO, NERC-BODC and ICES

SeaDataNet





1.87 million CDI entries from **34** countries, **102** data centres and **597** originators for physics, chemistry, geology, geophysics, bathymetry and biology; from **1805 to 2016**; **86%** unrestricted or under SDN License



- Build FCM (High and low resolution) standard common vocabularies
- Set up a common FCM data management protocols and methods
- 3. Connect FCM datasets to SDC infrastructure

Steps1: Build standard vocabularies



→ Set up a captured parameters for HR FCM



Steps1: Build standard vocabularies



→ Set up a captured parameters for HR FCM



Steps1: Build standard vocabularies



70

		Common Metadata	<mark>-</mark> L	Inique Metadata	Comm	on Data	Unique data
Rijkswaterstaa	t						
Université Litional Cole d'Opale							
VLIZ							
Institut Méditerranéen d'Océanologie							
	0	10 20)	30 4	0	50	60

→ 73 Captured parameters (Metadata+Data)



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Captured PARAM
Project
Project starting Date
Project ending Date
Person of Interset
Cytometer ID
Beam width
Core speed
nstrument name
CytoUSB Block size
Quality check instrument
lemperature QC (instrument
noise signal QC (instrument)
itation
Depth
.atitude
.ongitude
itudy area
amples Operator
itandards Reference
Clustering Method
Observation Type
Platform Type
Platform ID
Platform nationality
ampling Date
Analysis Date
Measurement duration
file version
low rate (μL/sec)
ile name
rigger Channel
rigger level
amplification#
las curvature
imart trigger
mart trigger level

	Standardized name							
	Selection Set							
	Volume							
	Abundance (cell/cm-3)							
	Particule number							
	Mean tot FWS							
	SD tot FWS							
	Mean tot SWS							
	SD tot SWS							
	Mean tot FLR							
	SD tot FLR							
	Mean tot FLO							
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Η.	Length FWS							
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	Tot_FWS							
	Tot_SWS-hs							
	Tot_SWS-Is							
	Tot_FLY -hs							
	Tot_FLO-hs							
	Total FL Red HS/ml							
	MaxFWS							
	Max SWS - hs							
	Max SWS - Is							
	Max FLY - hs							
	Max FLO - hs							
	Max FLR - hs							
	TOF							
	Beta 0 (coef intecept)							
	Beta 1 (coef (slope)							
	Mean Length							
	SD Length							
	link nic							



Some propositions for standardization

1. Provide an average cytograms/region and techniques (2 or 3) with these combinations such as: Tot FLR/ Tot FWS or/and Tot FLR/Tot FLO

Tot FLR

→ Determine the cytogram of a BLANCO sample (using sea water or sheath fluid) with identical trigger for phyto. analysis

➔ Determine the average cytogram « conceptual cyto » of the phytoplankton groups in (arbitary unit/cell)

Ecart-type du'Total de la fluorescence orange(FLO)'(u.a./ceil)



noise distinction

Tot FWS



2. Define phytoplankton groups in the point of view of the Cytometrist

SELECTION SET	STANDARDIZED NAMES	DEFINITION
Cryptophytes	Cryptophytes	A group characterized by an orange fluorescences cells with similar FWS as Nanoplancton. The specific definition depends on the laser wave bands. i.e: red laser will also excite phycocyanine. The Cryptophytes are also Nanoeukaryotes
Microphytoplankton	Microphytoplankton	A group where the cluster is above Nanoeukaryotes based on average cytogram
largephyto3 - N/mL	Microphytoplankton	
largephyt2 - N/mL	Microphytoplankton	
largephyto1 - N/mL	Microphytoplankton	
Nanoeukaryotes 1	Nanoeukaryotes	It is a group above Picoeukaryotes in terms of FWS and FLR
Nanoeukaryote 2s	Nanoeukaryotes	



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3. Check the 34 existing codes (NERC vocabulary server)

			,			
conceptid	id preflabel		altlabel	definition	Deprecate Y/N	Alternative code
PYPKAFB1	Abundance of Bacteria (ITIS: 202421: WoRMS 6) [Subgroup: group PSB1 autotrophic] per unit volume of the water body by flow cytometry	4/17/2016 15:50:07	Bact_PSB1_auto	Number of particles resolved as photosynthetic bacteria cells from the uncharacterised cluster PSB1 in a unit volume of any body of fresh or salt water determined by flow cytometry analysis of unstained samples.		
PYPKAFB2	Abundance of Bacteria (ITIS: 202421: WoRMS 6) [Subgroup: group PSB2 autotrophic] per unit volume of the water body by flow cytometry	4/17/2016 15:50:07	Bact_PSB2_auto	Number of particles resolved as photosynthetic bacteria cells from the uncharacterised cluster PSB2 in a unit volume of any body of fresh or salt water determined by flow cytometry analysis of unstained samples.		
P18318A9	Abundance of Bacteria (ITIS: 202421: WoRMS 6) [Subgroup: heterotrophic; high nucleic acid cell content] per unit volume of the water body by flow cytometry	4/17/2016 15:50:07	Abund_BE006316	Number of particles resolved as heterotrophic bacteria cells from the high nucleic acid content cluster (HNA) in a unit volume of any body of fresh or salt water determined by flow cytometry analysis of samples stained with a nucleic acid- specific fluorescent dye, and subtraction of cyanobacteria cell count if present.		
C804B6A6	Abundance of Bacteria (ITIS: 202421: WoRMS 6) [Subgroup: heterotrophic; low nucleic acid cell content] per unit volume of the water body by flow cytometry	4/17/2016 15:50:07	Abund_BE006317	Number of particles resolved as heterotrophic bacteria cells from the low nucleic acid content cluster (LNA) in a unit volume of any body of fresh or salt water determined by flow cytometry analysis of samples stained with a nucleic acid-specific fluorescent dye, and subtraction of cyanobacteria cell count if present.		

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P01 (BODC Parameter Usage Vocabulary) - existing vocabulary for flow cytometry

Conclusion



- MIO has been deploying Cytobuoy instruments for the last 10 years that allowed us to work on data management
- FCM processing tools have various output structures and formats → There is a need to build a conversion tool. Mathilde's tool is a good example and can be developed to be more common.
- We need to check the 34 existing FCM standardized codes.
- 73 Captured parameters (VLIZ, MIO,RWS and ULCO) → An urgent need to focus on functional group names definition : An average cytogram
- Standardizing FCM vocabulary will increase the visibility on EU plateform (SeaDataNet infrastructure) and join other datasets to make your data usefull to the community.

Thank you for building together FCM standards



Aix Marseille

Discussion Group1: Harmonisation of FCM use and data

- The classification of functional group: Prochlorococcus, Picoeukaryotes, Synechococcus, coccolitophore, Cryptophytes, Nanophytoplankton, Microphytoplankton, Dinoflagellates, Cyanobacteria.
- Definition of group: There is no need for that because it is already done. Also, because of the different clustering and optical properties. For the moment, we can't define the group on size criteria (since it is not clear yet).
- Clustering defnition and method depend on the operator if he is using manual or auto. Thus, the intercomparaison exercice within the JERICO Next to determine the discrepancy of the output of the clustering by manual/Auto.
- Standardization of the protocol depends on regions
- Standardization of sensor by beads and by chlorophyll
- Existing stand. FCM PARAM (NERC): some are good but there is a lot of redundancy and definition are not clear. Very difficult to understand and not helpful at all.
- Another forum to discuss about standardization.