Marseille, from 19 to 22 March 2018

JERICO next

JOINT EUROPEAN RESEARCH INFRASTRUCTURE NETWORK FOR COASTAL OBSERVATORIES

The setting up of new flow cytometry standardized and common vocabulary

SeaDataCloud

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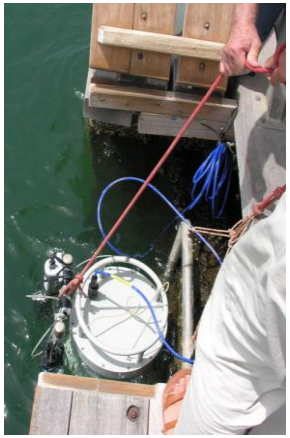
mio Institut Méditerranéen d'Océanologie | BODC | Institut Pythéas Observatoire des Sciences de l'Univers Aix*Marseille Université | Ifremer | MARIS

Third JERICO-NEXT Workshop on Phytoplankton Automated Observation/ MIO/ Marseille/ France/ 19th to 21st March 2018

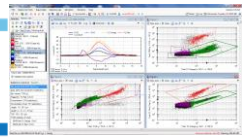
Introduction



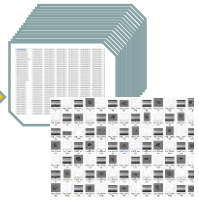
FCM Tools

VOCABULARY HETEROGENEITY
(Parameters, Cluster names, Unit, etc.)

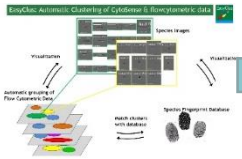
COMMON VOCABULARY



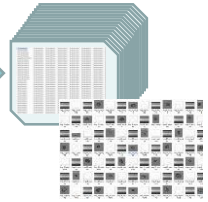
CytoClus



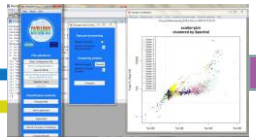
VOCAB 1



EasyClus



VOCAB 2



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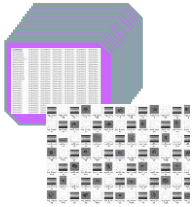

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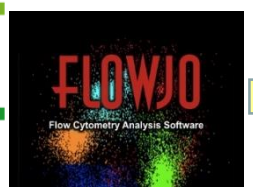
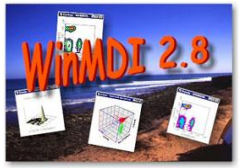
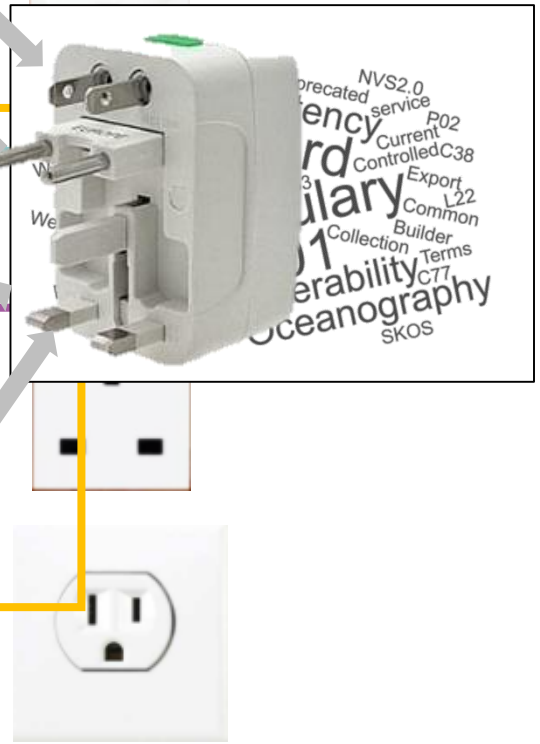


VOCAB n





FCM Tools

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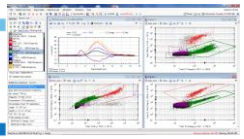
COMMON VOCABULARY



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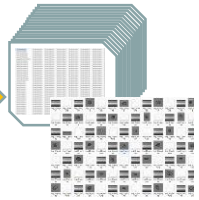
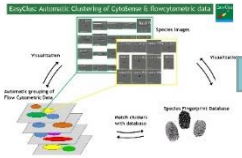
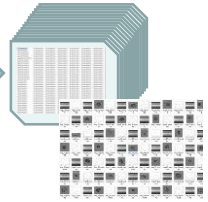


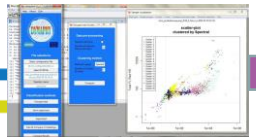
VOCAB 1



EasyClus



VOCAB 2



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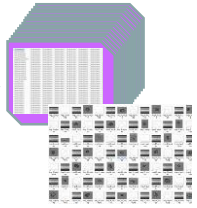
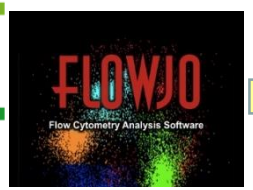
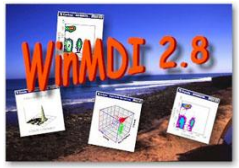
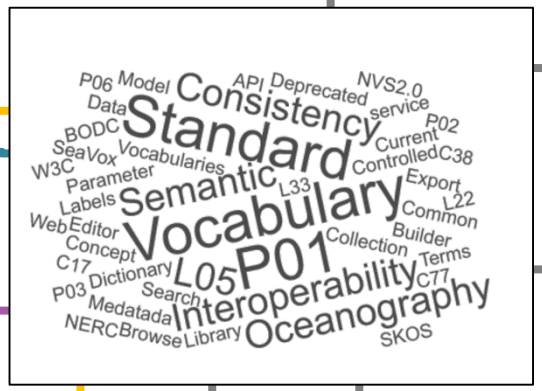


VOCAB 3



VOCAB n





SeaDataNet

EMODnet

European Marine Observation and Data Network

OBIS

OCEAN BIOGEOGRAPHIC INFORMATION SYSTEM

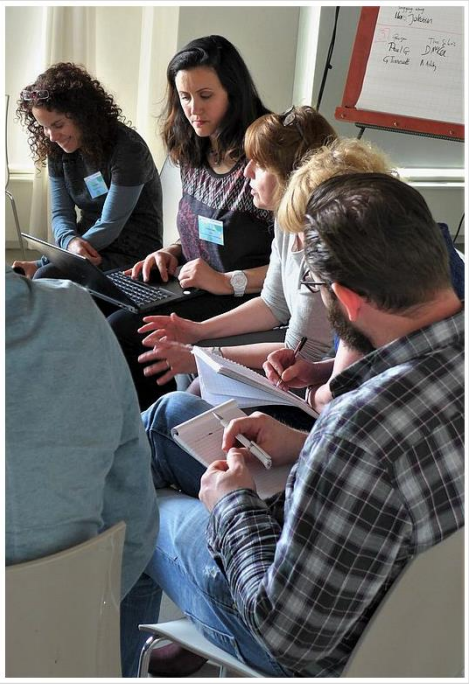
Other ISO Portals

FCM Common Vocabulary

1. Analysis of the existing FCM Common vocabulary
2. Captured parameters (Cefas, MIO, RWS, ULCO and VLIZ)
3. Setting up FCM Standardized common vocabulary
4. Questionnaire

1. Analysis of the existing Common vocabulary

Cytobuoy Meeting, March 2017➔ Parallel session – Harmonisation of flow cytometry use and data (protocol, standardisation, definition of functional types, quality control)



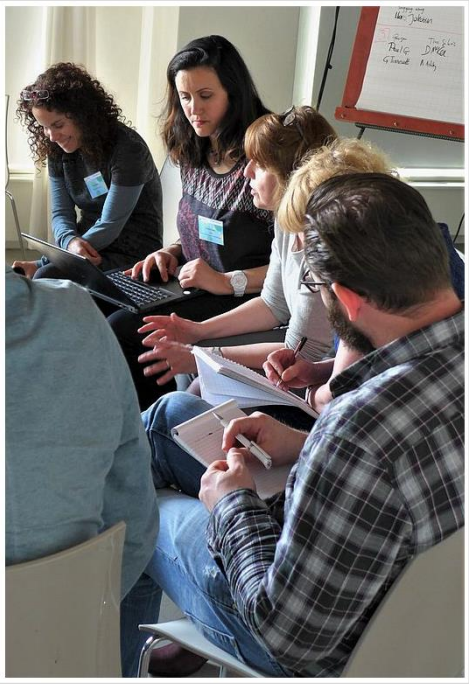
➔ 34 FCM codes existed in the BODC VS

P01 (BODC Parameter Usage Vocabulary) -existing vocabulary for flow cytometry

conceptid	preflabel	modified	altlabel	definition	Deprecate Y/N	FCM Community Feedback
PYPKAFB1	Abundance of Bacteria (ITIS: 202421: WoRMS 6) [Subgroup: group PSB1 autotrophic] per unit volume of the water body by flow cytometry	4/17/2016 15:50:07	Bact_PSB1_auto	Number of particles resolved as photosynthetic bacteria cells from the uncharacterised cluster PSB1 in a unit volume of any body of fresh or salt water determined by flow cytometry analysis of unstained samples.		Prochlorochocus they don't know what it is. It is not clear we need to separate syno and proclo and give definition for each one: difference by size and pigment.
PYPKAFB2	Abundance of Bacteria (ITIS: 202421: WoRMS 6) [Subgroup: group PSB2 autotrophic] per unit volume of the water body by flow cytometry	4/17/2016 15:50:07	Bact_PSB2_auto	Number of particles resolved as photosynthetic bacteria cells from the uncharacterised cluster PSB2 in a unit volume of any body of fresh or salt water determined by flow cytometry analysis of unstained samples.		Synechococcus (1 - 2 um)
P18318A9	Abundance of Bacteria (ITIS: 202421: WoRMS 6) [Subgroup: heterotrophic; high nucleic acid cell content] per unit volume of the water body by flow cytometry	4/17/2016 15:50:07	Abund_BE006316	Number of particles resolved as heterotrophic bacteria cells from the high nucleic acid content cluster (HNA) in a unit volume of any body of fresh or salt water determined by flow cytometry analysis of samples stained with a nucleic acid-specific fluorescent dye, and subtraction of cyanobacteria cell count if present.		Not concerned
ADI07845	Abundance of Dinoflagellates (ITIS: 9873: WoRMS 19542) [Size: <20um Subgroup: autotrophic] per unit volume of the water body by flow cytometry	2/22/2016 14:44:37	ADino<20um_FC	Number of particles <20um identified as most likely small dinoflagellates in a unit volume of any body of fresh or salt water determined by flow cytometry analysis of unstained samples.		How can you define them by FCM??
P200A00Z	Abundance of Dinoflagellates (ITIS: 9873: WoRMS 19542) per unit volume of the water body by flow cytometry	2/18/2016 15:18:01	P200A00Z	This code was updated on 18-feb-2016 so that the name Dinoflagellates is used instead of Pyrrophycophyta in the preferred label field. The vernacular term 'dinoflagellates' maps to Pyrrophycophyta in the ITIS taxonomy and Dinophyceae in the WoRMS taxonomy. In the		not agree and not clear
PU00A02A	Abundance of eukaryote picophytoplankton per unit volume of the water body by flow cytometry	4/17/2016 15:50:07	UndifEuPicoPhytAbund	Number of particles resolved as photosynthetic eukaryote cells in the picoplankton size range in a unit volume of any body of fresh or salt water determined by flow cytometry analysis of unstained samples.		not agree and not clear

1. Analysis of the existing Common vocabulary

Cytobuoy Meeting, March 2017➔ Parallel session – Harmonisation of flow cytometry use and data (protocol, standardisation, definition of functional types, quality control)

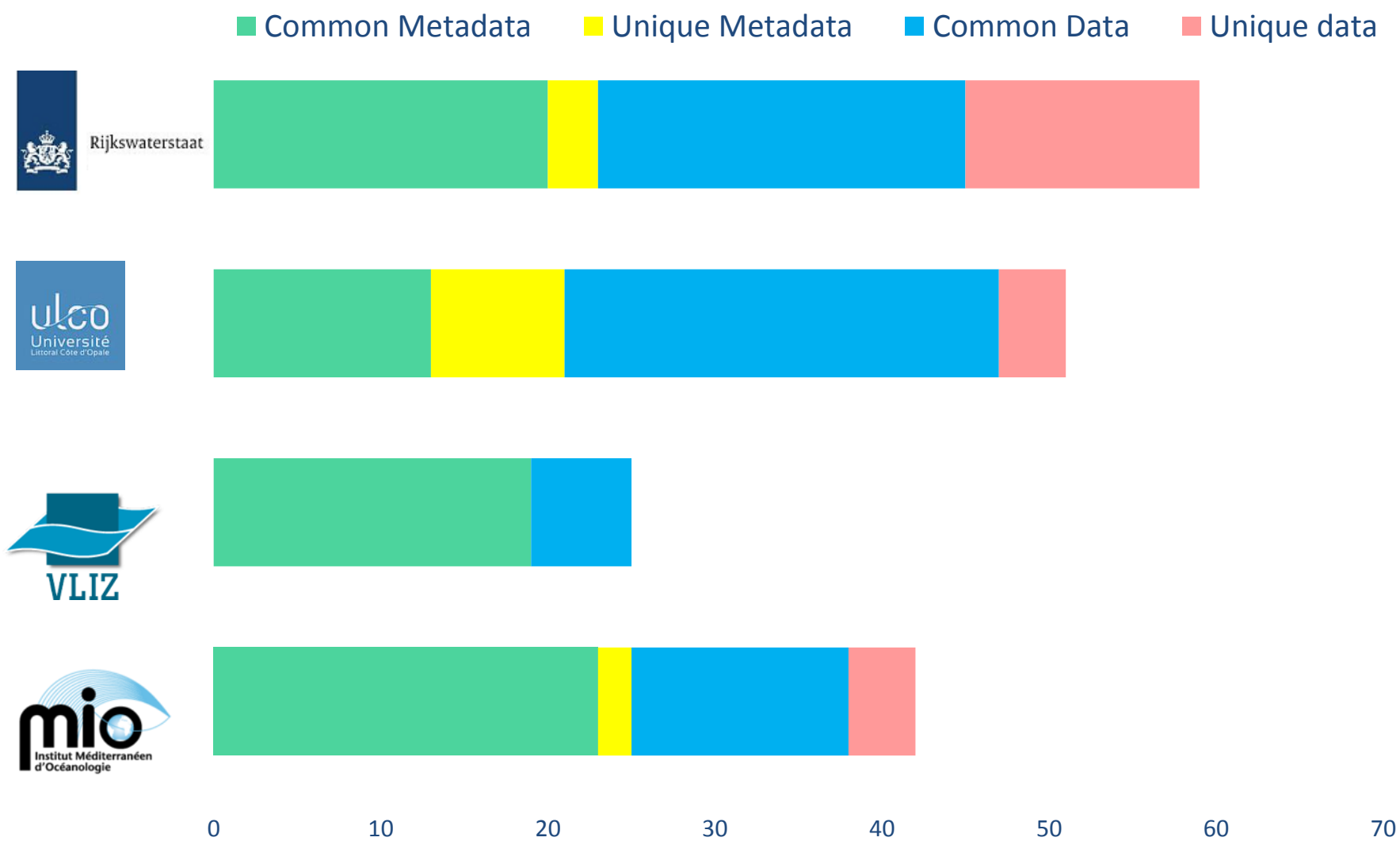


➔ 34 FCM codes existed in the BODC VS

- Some codes are good
- Redundancy
- Definitions are not clear for FCM users and difficult to understand

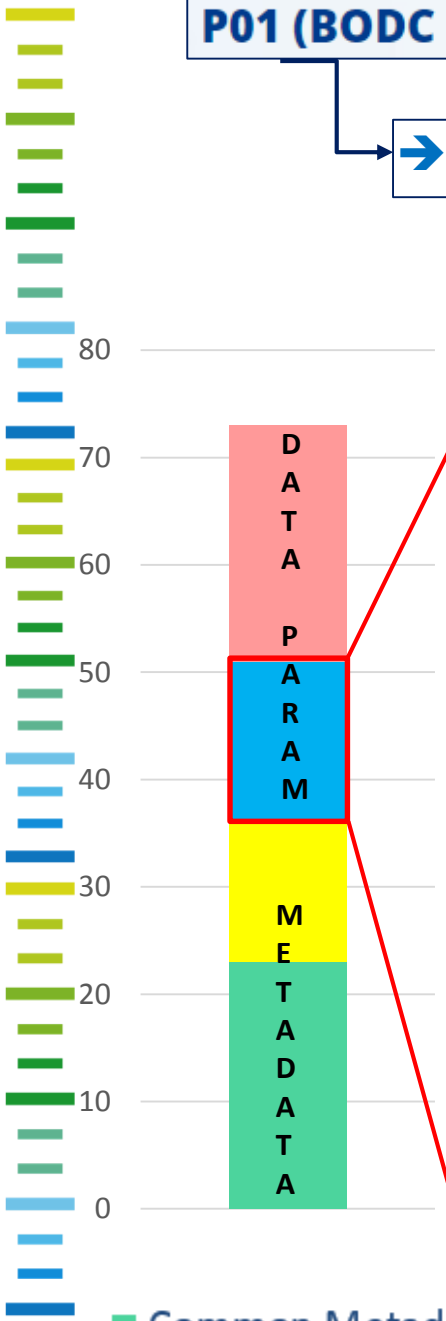
➔ Build a new FCM Common vocabulary based on the measured quantities

2. FCM captured parameters



P01 (BODC PARAMETER USAGE VOCABULARY)

➔ 24 Captured Common Data paramters sent to BODC



12 Common Param. (Area)

- FWS_average_**area**
- FWS_SD_**area**
- SWS_average_**area**
- SWS_SD_**area**
- FLR_average_**area**
- FLR_SD_**area**
- FLO_average_**area**
- FLO_SD_**area**
- FLY_average_**area**
- FLY_SD_**area**
- FLG_average_**area**
- FLG_SD_**area**

12 Common Param. (Height/Max)

- FWS_average_**height**
- FWS_SD_**height**
- FWS_average_**height**
- FWS_SD_**height**
- FLR_average_**height**
- FLR_SD_**height**
- FLO_average_**height**
- FLO_SD_**height**
- FLY_average_**height**
- FLY_SD_**height**
- FLG_average_**height**
- FLG_SD_**height**

Common Metadata Unique Metadata Common Data Unique data

3. Setting the FCM Standardized Common Vocabulary

- Semantic model (BODC)

Chemical model	Biological model	Physical model
Measurement Substance Measurement matrix relationship Matrix Matrix subcomponent	Measurement Organism Name Organism Specifics Measurement matrix relationship Matrix Matrix subcomponent Method	Measurement Statistical Physical entity Measurement matrix relationship Matrix Method
Concentration of carbon (total inorganic) {TCO2} per unit mass of the water body [dissolved plus reactive particulate phase]	**Abundance** of Bacteria (ITIS: 202421: WoRMS 6) [Subgroup: heterotrophic] per unit volume of the water body by automated flow cytometry	**Area mean of** Forward light scatter pulse per cluster from the water body by flow cytometry

Area

Mean of

Forward light scatter pulse

per cluster from the

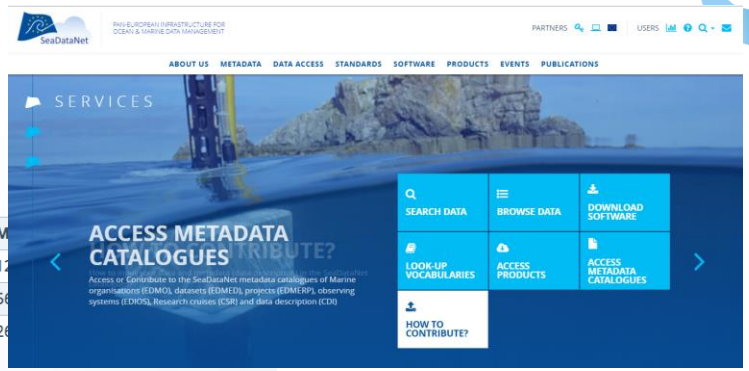
Water body by

flow cytometry

The cluster name is managed in a separate vocabulary list

- FCM Standardized Common Vocabulary

➔ https://www.seadatanet.org/



BODC WEBSERVICES V2 (LIBRARIES) CL12

Library	Thesaurus	Title	Alt Title	Version	M	
C16		SeaDataNet sea areas	SDN sea areas	9	12	
C17		ICES Platform Codes	ICES Platforms	712	56	
C19		SeaVoX salt and fresh water body gazetteer	SeaVoX water bodies	17	26	
C32		International Standards Organisation countries	ISO countries	7	251	1/14/2016 2:00:02 AM
C34		Activity purpose categories	Purpose categories	4	22	8/27/2011 3:00:05 AM
C35		European Nature Information System	EUNIS3 Habitats	1	56	2/19/2010 2:01:37 AM

F02	SeaDataCloud Flow Cytometry Standardised Cluster Names	SDC flow cytometry cluster names	2	11	2/3/2018 2:00:02 AM
P01	BODC Parameter Usage Vocabulary	BODC PUV	800	37732	3/14/2018 2:00:03 AM
P02	SeaDataNet Parameter Discovery Vocabulary	SeaDataNet PDV	107	435	2/13/2018 2:00:03 AM
L22	SeaVoX Device Catalogue	SeaVoX Device Catalogue	324	1280	3/6/2018 2:00:04 AM
P06	BODC data storage units	BODC units	99	346	2/16/2018 2:00:02 AM

F02 (SEADATACLOUD FLOW CYTOMETRY STANDARDISED CLUSTER NAMES)

ConceptID	Preferred label	Alt label	Definition	Modified
F0200001	Standard beads		A standard is a reference defined by a user, a laboratory, or any acknowledged authority. Properties of standard beads are accurately known by the manufacturers (i.e. size, material, fluorescence properties). These fluorescent microbeads (or microsphere) are used as an absolute reference for quantitative and qualitative comparisons. Standard beads are analyzed routinely in every flow cytometry analyses in order to have confidence in the instrument performance (alignment and fluidics) and as well as in the results.	11/20/2017 13:09:10
F0200002	Prochlorococcus		Prochlorococcus cells are defined as the smallest cyanobacteria found in the marine environment. No staining is required to distinguish them by flow cytometry. Compared to any other group, their forward scatter and red fluorescence signatures are the smallest recorded up to now and require sensitive Photo Multiplier Tube (PMT) or high powered lasers. The cluster, when well defined (often deep water communities) is below or may overlap that of the Synechococcus group, and is	11/20/2017 13:09:10

P01 (BODC PARAMETER USAGE VOCABULARY)

Overview | Export subset of list | Export full list | New query | Found 2 | Current | Previous | Next

ConceptID	Preferred label	Alt label	Definition	Modified
SCNAME01	Taxon of biological entity specified elsewhere	Name_BE007117	The scientific name of the biological object.	1/21/2016 13:55:16
SNANID01	Identifier (LSID) of biological entity specified elsewhere	LSID_BE007117	A global unique identifier for the nomenclatural details of the scientific name of a biological object (urn:lsid:marinespecies.org:taxname:ID)	1/21/2016 13:55:16

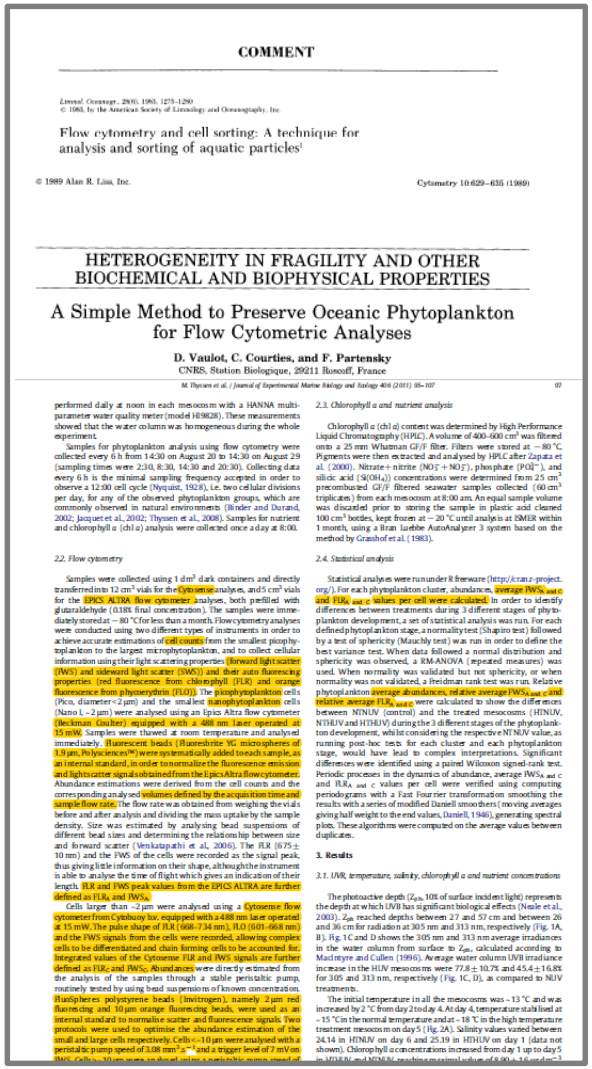
Example: SCNAME01= Akashiwo sanguinea and SNANID01= urn:lsid:marinespecies.org:taxname:232546

ConceptID	Preferred label	Alt label	Definition	Modified
NMCLFL02	Registered name of flow cytometry cluster by classification to a term from the NVS SeaDataCloud Flow Cytometry Standardised Cluster Names Vocabulary (SDN:F02::)	ClusterName	Text term identifying the type of particles belonging to a specific flow cytometry cluster, taken from the NVS SeaDataCloud Flow Cytometry Standardised Cluster Names controlled vocabulary F02.	2/1/2018 21:53:44
IDCLFL02	Registered name identifier of flow cytometry cluster by classification to a term from the NVS SeaDataCloud Flow Cytometry Standardised Cluster Names Vocabulary (SDN:F02::)	ClusterNameID	Opaque key term identifying the type of particles belonging to a specific flow cytometry cluster, taken from the NVS SeaDataCloud Flow Cytometry Standardised Cluster Names controlled vocabulary F02	2/1/2018 21:53:44

Example: NMCLFL02 = Eukaryote nanophytoplankton and IDCLFL02 = SDN:F02::F0200005

3. FCM Questionnaire

- Literrature review from 1983 till 2017



COMMENT

Limnol. Oceanogr., 28(6), 1983, 1273–1280
© 1983, by the American Society of Limnology and Oceanography, Inc.

Flow cytometry and cell sorting: A technique for analysis and sorting of aquatic particles[1]

© 1989 Alan R. Liss, Inc. Cytometry 10:629–635 (1989)

HETEROGENEITY IN FRAGILITY AND OTHER BIOCHEMICAL AND BIOPHYSICAL PROPERTIES

A Simple Method to Preserve Oceanic Phytoplankton for Flow Cytometric Analyses

D. Vaulot, C. Courties, and F. Partensky
CNRS, Station Biologique, 29211 Roscoff, France

M. Thyssen et al. / Journal of Experimental Marine Biology and Ecology 406 (2011) 95–107

performed daily at noon in each mesocosm with a HANNA multi-parameter water quality meter (model H98828). These measurements showed that the water column was homogeneous during the whole experiment.

Samples for phytoplankton analysis using flow cytometry were collected every 6 h from 14:30 on August 20 to 14:30 on August 29 (sampling times were 2:30, 8:30, 14:30 and 20:30). Collecting data every 6 h is the minimal sampling frequency accepted in order to observe a 12:00 cell cycle (Nyquist, 1928), i.e. two cellular divisions per day, for any of the observed phytoplankton groups, which are commonly observed in natural environments (Binder and Durand, 2002; Jacquet et al., 2002; Thyssen et al., 2008). Samples for nutrient and chlorophyll *a* (chl *a*) analysis were collected once a day at 8:00.

2.2. Flow cytometry

Samples were collected using 1 dm³ dark containers and directly transferred into 12 cm³ vials for the Cytosense analyses, and 5 cm³ vials for the EPICS ALTRA flow cytometer analyses, both prefilled with glutaraldehyde (0.18% final concentration). The samples were immediately stored at −80 °C for less than a month. Flow cytometry analyses were conducted using two different types of instruments in order to achieve accurate estimations of cell counts from the smallest picophytoplankton to the largest microphytoplankton, and to collect cellular information using their light scattering properties (forward light scatter (FWS) and sideward light scatter (SWS)) and their auto fluorescing properties (red fluorescence from chlorophyll (FLR) and orange fluorescence from phycoerythrin (FLO)). The picophytoplankton cells (Pico, diameter<2 μm) and the smallest nanophytoplankton cells (Nano, 1–2 μm) were analysed using an Epics Altra flow cytometer (Beckman Coulter) equipped with a 488 nm laser operated at 15 mW. Samples were thawed at room temperature and analysed immediately. Fluorescent beads (Fluoresbrite YG microspheres of 1.9 μm, Polysciences™) were systematically added to each sample, as an internal standard, in order to normalize the fluorescence emission and light scatter signals obtained from the Epics Altra flow cytometer. Abundance estimations were derived from the cell counts and the corresponding analysed volumes defined by the acquisition time and sample flow rate. The flow rate was obtained from weighing the vials before and after analysis and dividing the mass uptake by the sample density. Size was estimated by analysing bead suspensions of different bead sizes and determining the relationship between size and forward scatter (Venkatapathi et al., 2006). The FLR (675±10 nm) and the FWS of the cells were recorded as the signal peak, thus giving little information on their shape, although the instrument is able to analyse the time of flight which gives an indication of their length. FLR and FWS peak values from the EPICS ALTRA are further defined as FLR_A and FWS_A.

Cells larger than ~2 μm were analysed using a Cytosense flow cytometer from Cytobuoy bv, equipped with a 488 nm laser operated at 15 mW. The pulse shape of FLR (668–734 nm), FLO (601–668 nm) and the FWS signals from the cells were recorded, allowing complex cells to be differentiated and chain forming cells to be accounted for. Integrated values of the Cytosense FLR and FWS signals are further defined as FLR_C and FWS_C. Abundances were directly estimated from the analysis of the samples through a stable peristaltic pump, routinely tested by using bead suspensions of known concentration. FluoSpheres polystyrene beads (Invitrogen), namely 2 μm red fluorescing and 10 μm orange fluorescing beads, were used as an internal standard to normalize scatter and fluorescence signals. Two protocols were used to optimise the abundance estimation of the small and large cells respectively. Cells<~10 μm were analysed with a peristaltic pump speed of 3.08 mm³ s⁻¹ and a trigger level of 7 mV on [illegible]

2.3. Chlorophyll a and nutrient analysis

Chlorophyll *a* (chl *a*) content was determined by High Performance Liquid Chromatography (HPLC). A volume of 400–600 cm³ was filtered onto a 25 mm Whatman GF/F filter. Filters were stored at −80 °C. Pigments were then extracted and analysed by HPLC after Zapata et al. (2000). Nitrate+nitrite ($NO_3^- + NO_2^-$), phosphate (PO_4^{3-}), and silicic acid ($Si(OH)_4$) concentrations were determined from 25 cm³ preconbusted GF/F filtered seawater samples collected (60 cm³ triplicates) from each mesocosm at 8:00 am. An equal sample volume was discarded prior to storing the sample in plastic acid cleaned 100 cm³ bottles, kept frozen at −20 °C until analysis at ISMER within 1 month, using a Bran Luebbe AutoAnalyzer 3 system based on the method by Grasshof et al. (1983).

2.4. Statistical analysis

Statistical analyses were run under R freeware (http://cran.r-project.org/). For each phytoplankton cluster, abundances, average $FWS_{A\ and\ C}$ and $FLR_{C\ and\ A}$ values per cell were calculated. In order to identify differences between treatments during 3 different stages of phytoplankton development, a set of statistical analysis was run. For each defined phytoplankton stage, a normality test (Shapiro test) followed by a test of sphericity (Mauchly test) was run in order to define the best variance test. When data followed a normal distribution and sphericity was observed, a RM-ANOVA (repeated measures) was used. When normality was validated but not sphericity, or when normality was not validated, a Freidman rank test was run. Relative phytoplankton average abundances, relative average $FWS_{A\ and\ C}$ and relative average $FLR_{C\ and\ A}$ were calculated to show the differences between NTNUV (control) and the treated mesocosms (HTNUV, NTHUV and HTHUV) during the 3 different stages of the phytoplankton development, whilst considering the respective NTNUV value, as running post-hoc tests for each cluster and each phytoplankton stage, would have lead to complex interpretations. Significant differences were identified using a paired Wilcoxon signed-rank test. Periodic processes in the dynamics of abundance, average $FWS_{A\ and\ C}$ and $FLR_{A\ and\ C}$ values per cell were verified using computing periodograms with a Fast Fourier transformation smoothing the results with a series of modified Daniell smoothers (moving averages giving half weight to the end values, Daniell, 1946), generating spectral plots. These algorithms were computed on the average values between duplicates.

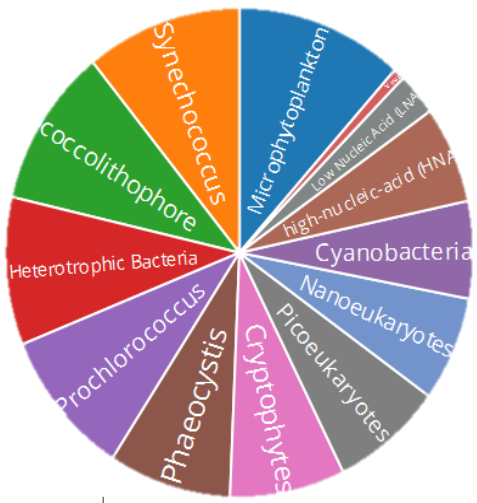
3. Results

3.1. UVR, temperature, salinity, chlorophyll a and nutrient concentrations

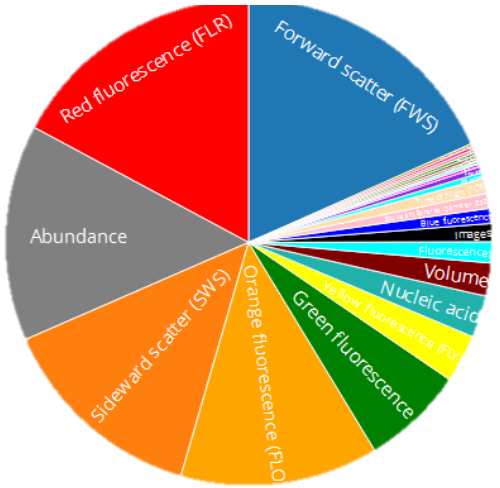
The photoactive depth (Z_{eu}, 10% of surface incident light) represents the depth at which UVB has significant biological effects (Neale et al., 2003). Z_{eu} reached depths between 27 and 57 cm and between 26 and 36 cm for radiation at 305 nm and 313 nm, respectively (Fig. 1A, B). Fig. 1C and D shows the 305 nm and 313 nm average irradiances in the water column from surface to Z_{eu}, calculated according to MacIntyre and Cullen (1996). Average water column UVB irradiance increase in the HUV mesocosms were 77.8±10.7% and 45.4±16.8% for 305 and 313 nm, respectively (Fig. 1C, D), as compared to NUV treatments.

The initial temperature in all the mesocosms was ~13 °C and was increased by 2 °C from day 2 to day 4. At day 4, temperature stabilised at ~15 °C in the normal temperature and at ~18 °C in the high temperature treatment mesocosms on day 5 (Fig. 2A). Salinity values varied between 24.14 in HTNUV on day 6 and 25.19 in HTHUV on day 1 (data not shown). Chlorophyll *a* concentrations increased from day 1 up to day 5 in HTHUV and NTNUV, reaching maximal values of 4.80 to 14 μg dm⁻³

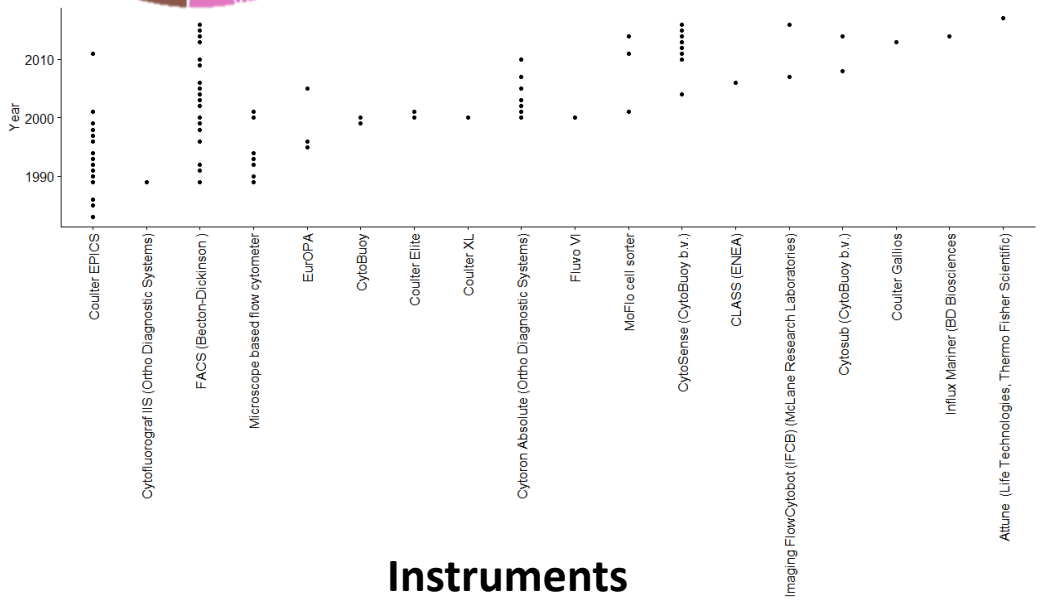
Clusters

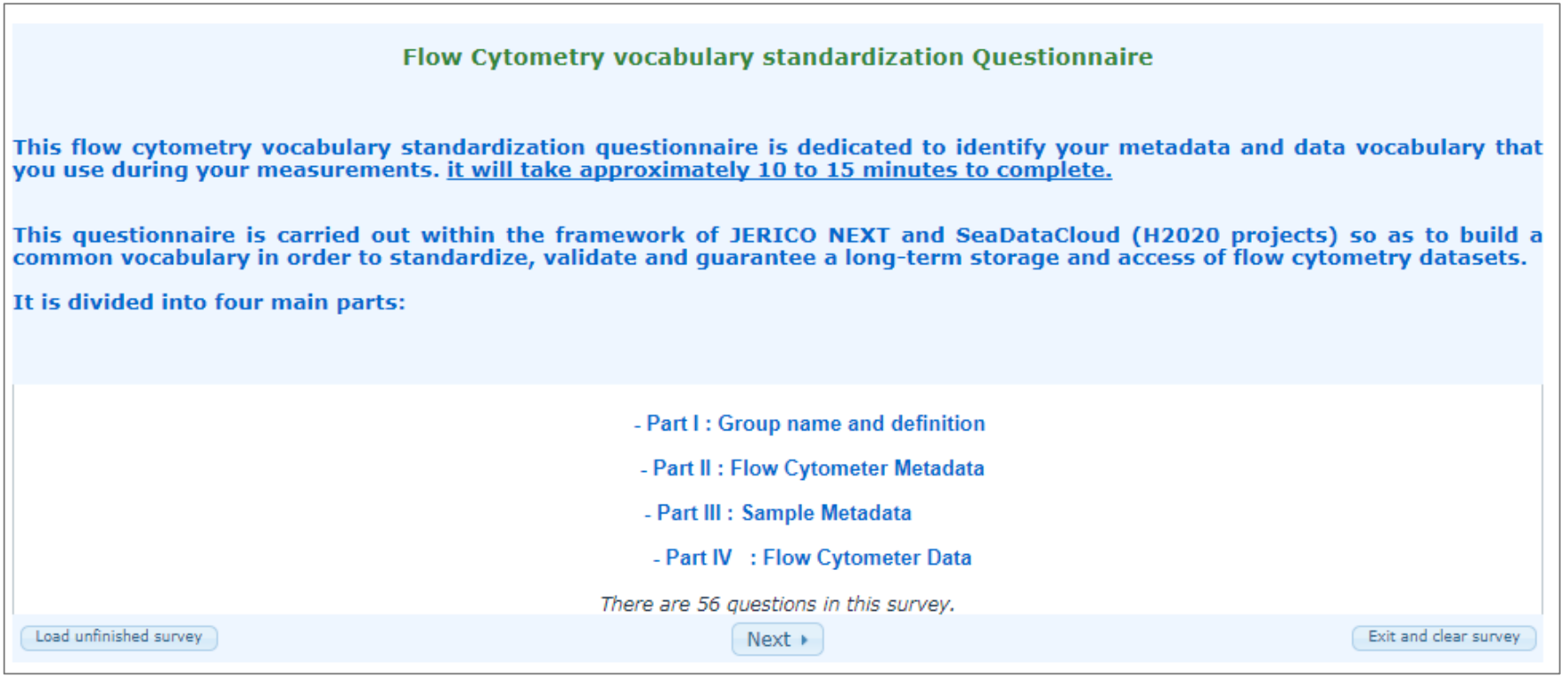


Measured parameters



Instruments





Flow Cytometry vocabulary standardization Questionnaire

This flow cytometry vocabulary standardization questionnaire is dedicated to identify your metadata and data vocabulary that you use during your measurements. it will take approximately 10 to 15 minutes to complete.

This questionnaire is carried out within the framework of JERICO NEXT and SeaDataCloud (H2020 projects) so as to build a common vocabulary in order to standardize, validate and guarantee a long-term storage and access of flow cytometry datasets.

It is divided into four main parts:

- Part I : Group name and definition
- Part II : Flow Cytometer Metadata
- Part III : Sample Metadata
- Part IV : Flow Cytometer Data

There are 56 questions in this survey.

Load unfinished survey | Next ▸ | Exit and clear survey

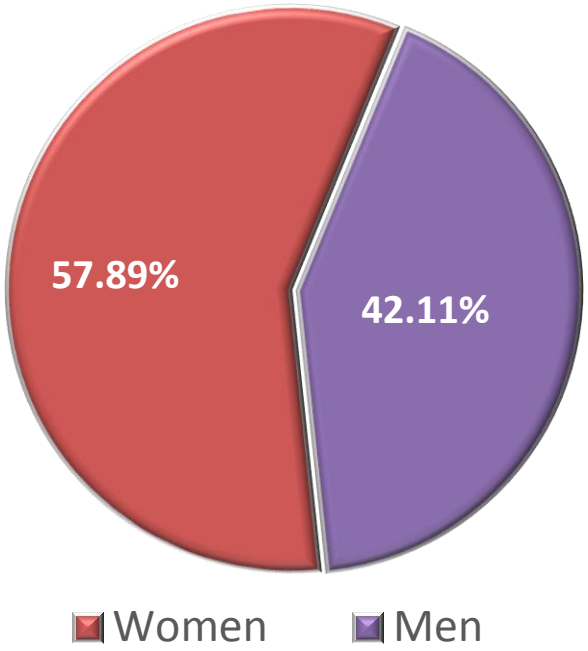
- Questionnaire sent to 180 FCM users all around the world

38 Answers to questionnaire from Oct. 30, 2017 to Jan. 15th 2018

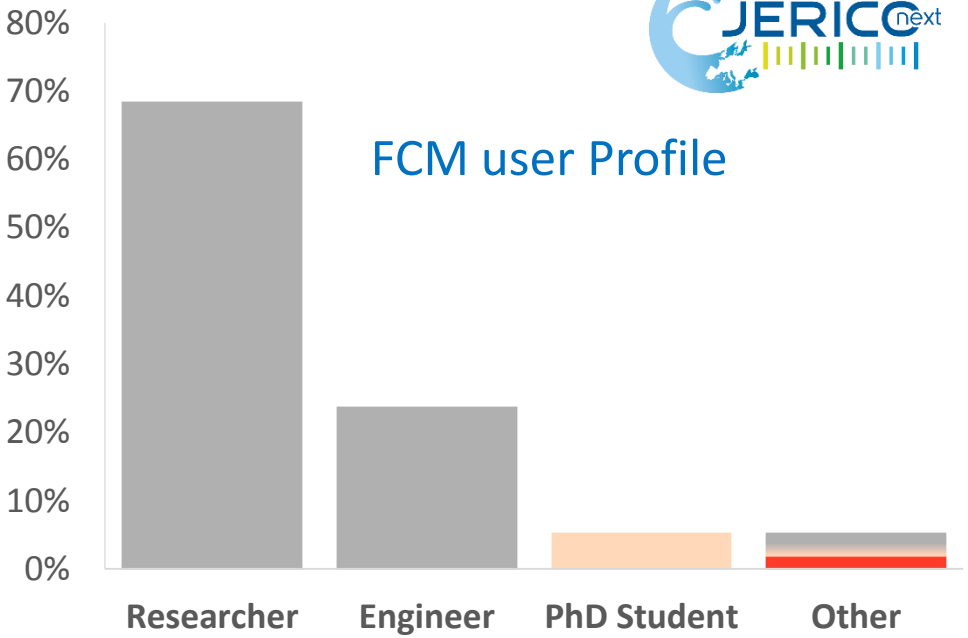
Uncompleted answers (21%)

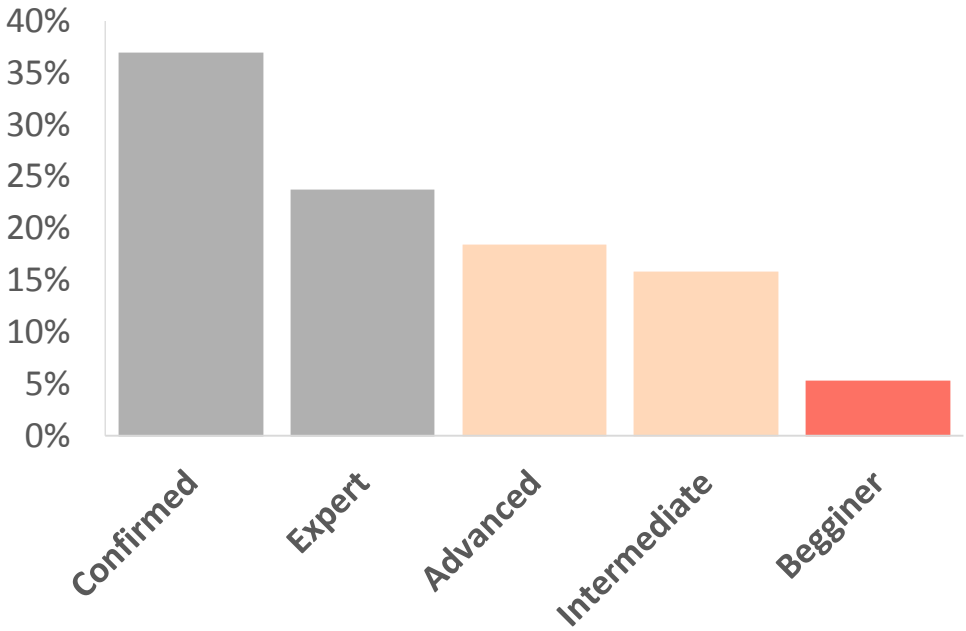
Completed answers (79%)

Gender participation



FCM user Profile

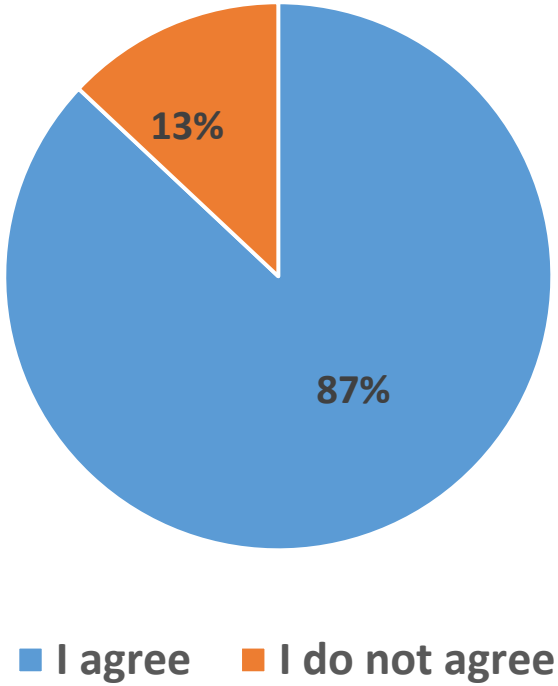




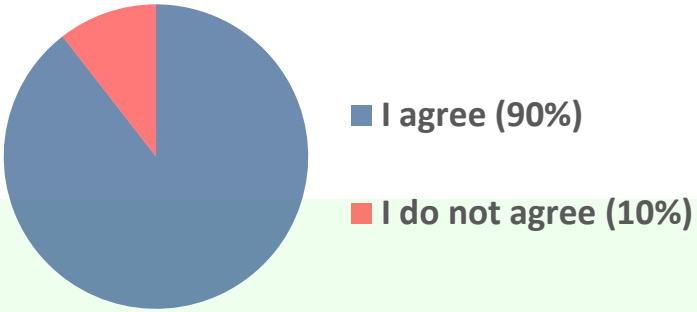
PART I: Groups definition from the FCM point of view

Q: Based on litterature from 1983 to 2017, do you agree on these group definitions:

Prochlorococcus, Synechococcus, Eukaryotes Picophytoplankton, Eukaryote Nanophytoplankton, Cryptophytes, Coccolithophores, Microphytoplankton and Heterotrophic Bacteria



Suggestion



* Prochlorococcus

Prochlorococcus are defined as the smallest cyanobacteria found in marine environment. No staining is required to distinguish them by flow cytometry. FWS and FLR signatures are the smallest recorded up to now and require sensitive PMT or high powered lasers. The cluster, when well defined (often deep water communities) is below or may overlap that of Synechococcus group, ~~and is often partially masked by the instrument background noise.~~

In samples stained for Heterotrophic bacteria analysis, Prochlorococcus can be distinguished using Sideward Scatter (SWS) vs Chlorophyll Red Fluorescences (FLR) cytogram. They do not emit orange fluorescence because they lack phycoerythrin.

* Prochlorococcus

Prochlorococcus are defined as the smallest cyanobacteria found in marine environment. No staining is required to distinguish them by flow cytometry. FWS and FLR signatures are the smallest recorded for any photosynthetic organisms up to now and require sensitive PMT or high powered lasers. The cluster, when well defined (often deep water communities) is below or may overlap that of Synechococcus group, and is NOT masked by the instrument background noise. When the cluster is not well defined, that is when it is partially masked by the noise.

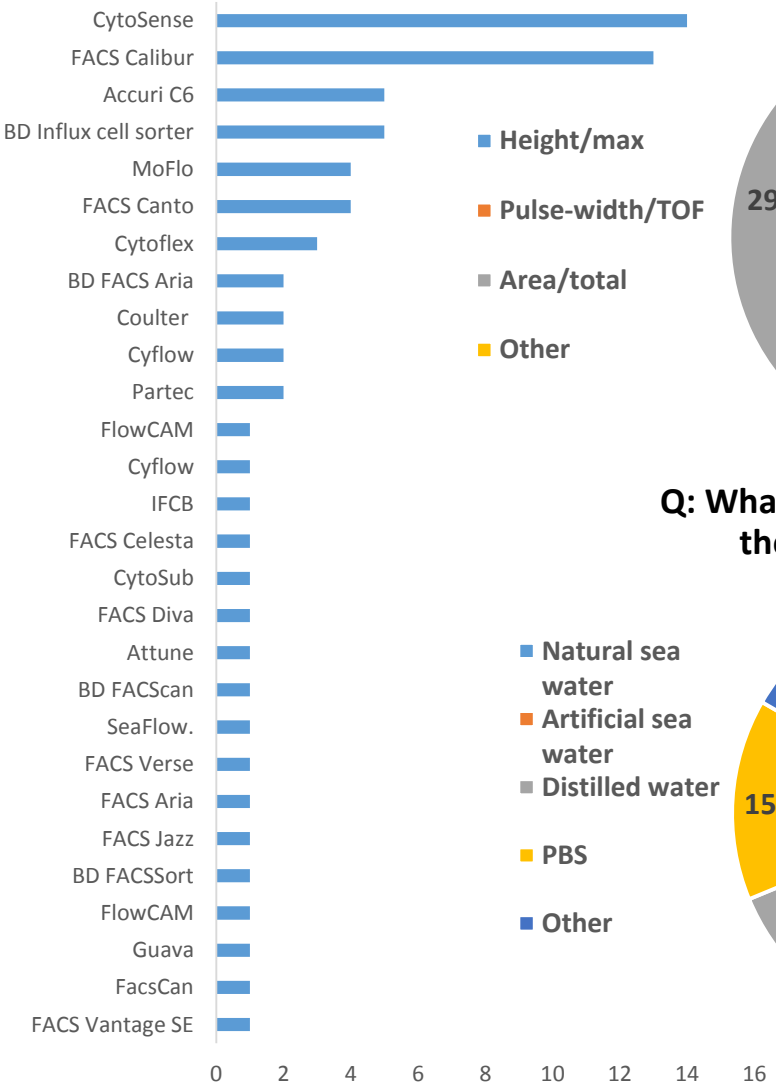
In samples stained for Heterotrophic bacteria analysis, Prochlorococcus can be distinguished, particularly deep communities (but not surface communities), using Sideward Scatter (SWS) vs Chlorophyll Red Fluorescences (FLR) cytogram. They do not emit orange fluorescence because they lack phycoerythrin.

NOTE:

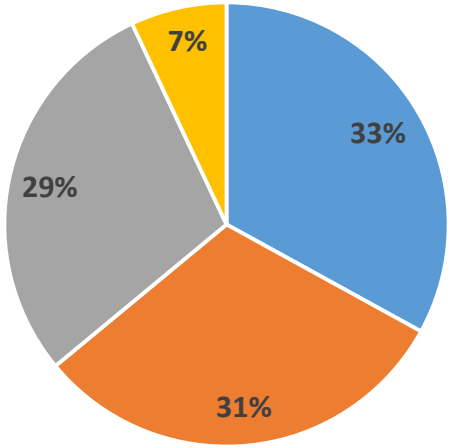
- *often the background noise is mostly NOT due to the instrument. It is due to heterotrophic bacteria, if the instrument is a good one.*

PART II: Flow Cytometer Metadata

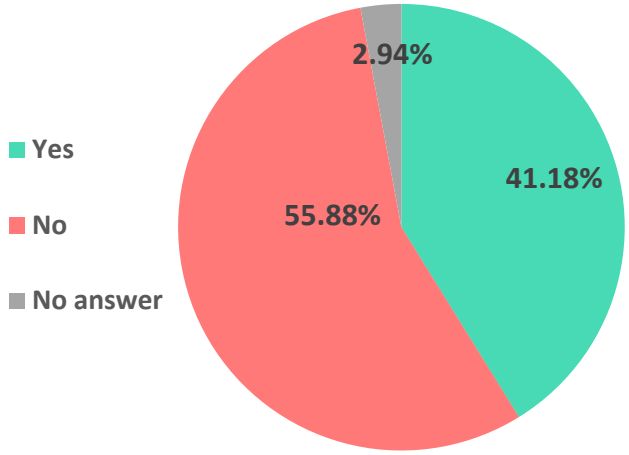
Q: What model of Flow Cytometer(s) do you use?



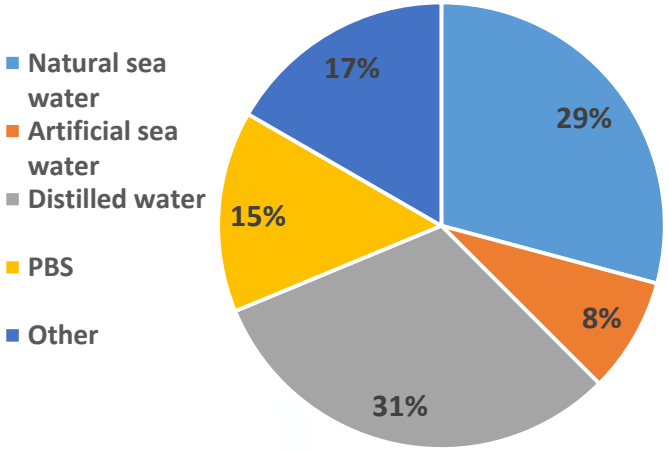
Q: What type of signal does your instrument record ?



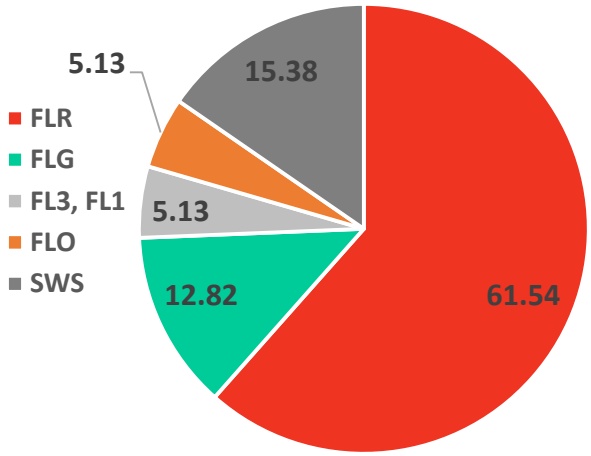
Q: Does your instrument have an image in flow device?



Q: What is the type/composition of the sheath fluid you use?

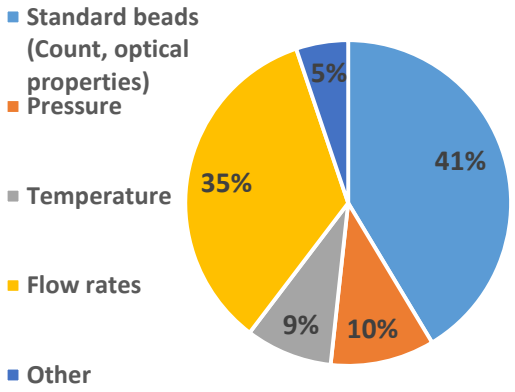


Q: What signal do you use as trigger?

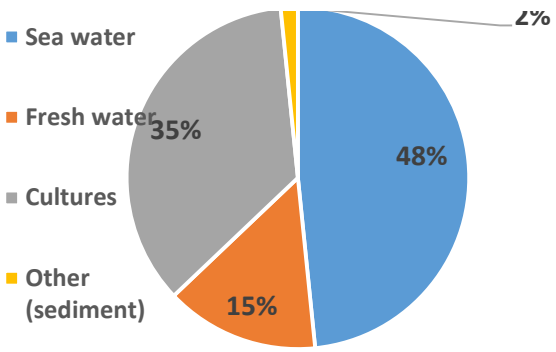


PART III: Sample Metadata

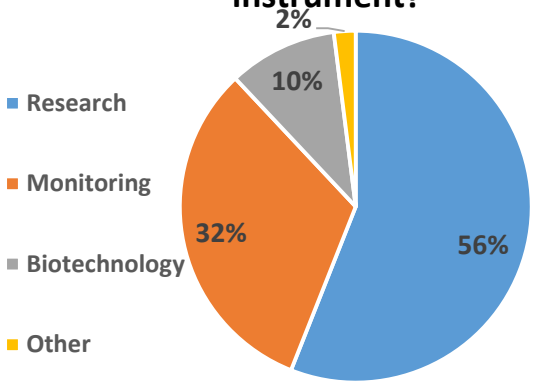
Q: How do you make the quality control



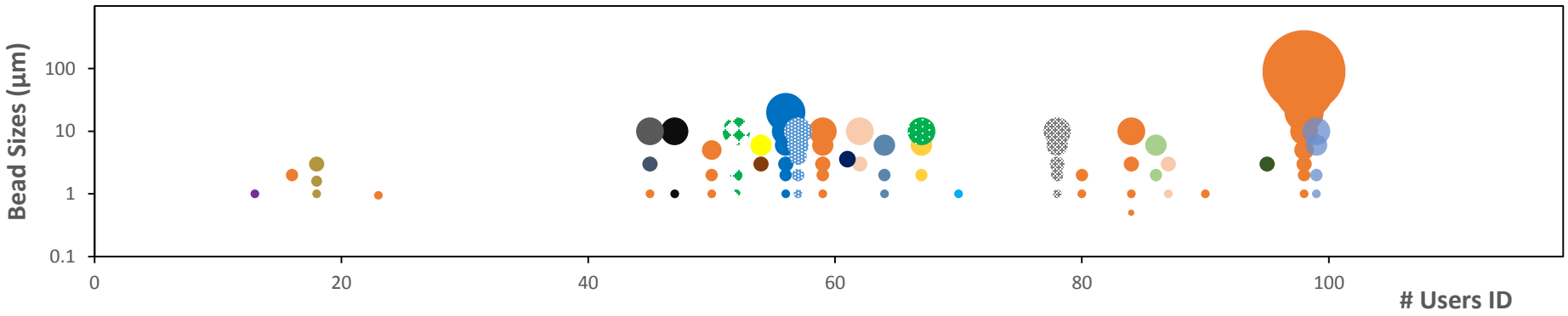
Q: What type of sample do you analyze?



Q: For what purpose do you use this instrument?



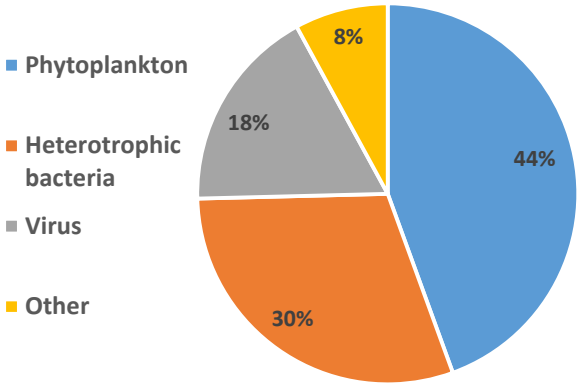
Q: What beads and diameters reference do you use ?



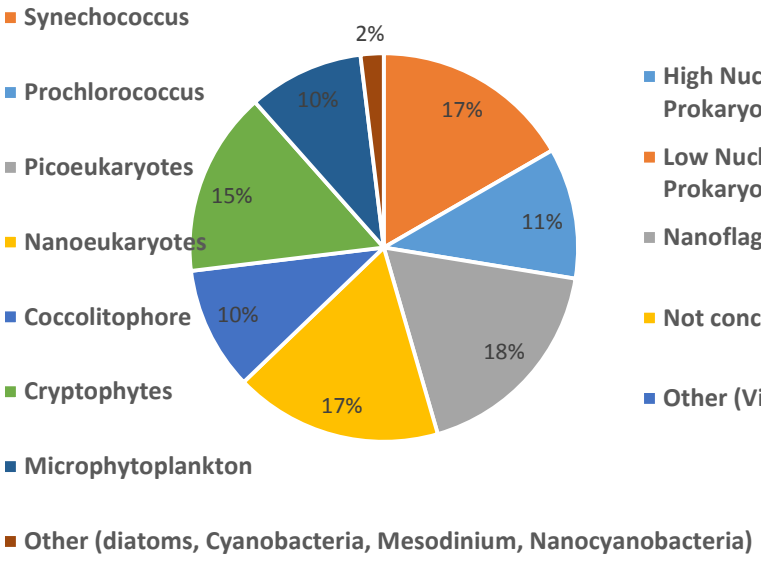
- Polyscience
- Thermofisher
- Beckman Coulter Flowset
- Calibration beads
- CountBrights
- Flow check high intensity beads
- Flow Check™ High Intensity
- Fluoresbrite Carboxy
- Fluorescence
- Fluorosphere
- FluoSpheres™ Polystyrene Microspheres
- FluoSpheres™ Sulfate Microspheres
- Invitrogen Fluospheres
- Molecular Probes
- Polysciences polybeads
- Polysciences, fluoresbrite YG microsphere
- Polystyrene microsphere
- Submicron beads
- Supplied by CytoBuoy
- ThermoFisher fluosphere
- Trucount Becton Dickinson

PART III: Sample Metadata

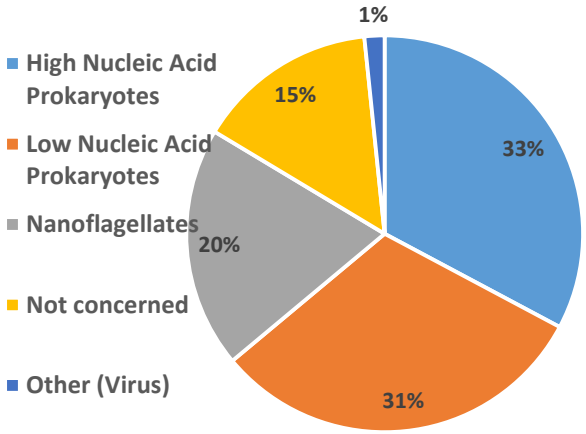
Q: Which type of particles do you measure?



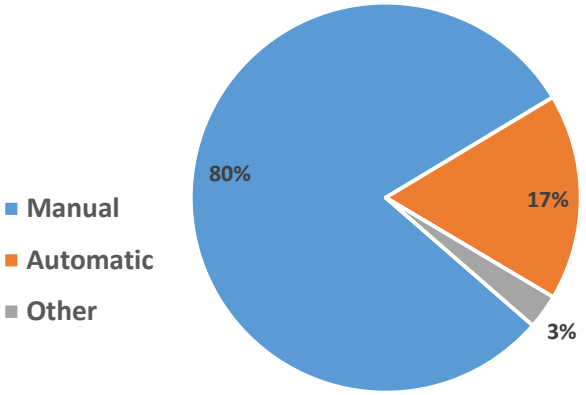
Q: What are the recurrent autotrophic functional groups of your area of study?



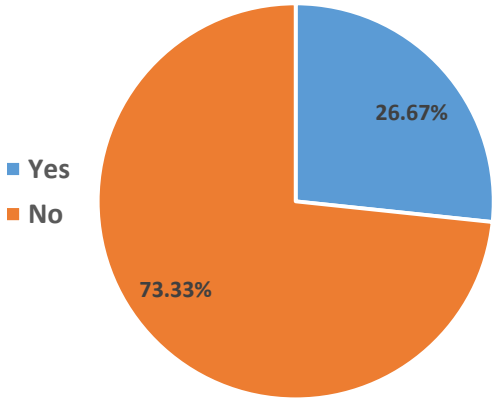
Q: What are the recurrent Heterotrophic groups of your area of study?



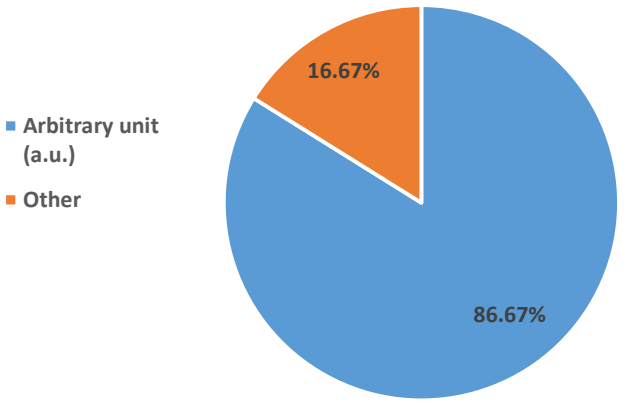
Q: Which clustering method do you use?



Q: Do you flag your data ?

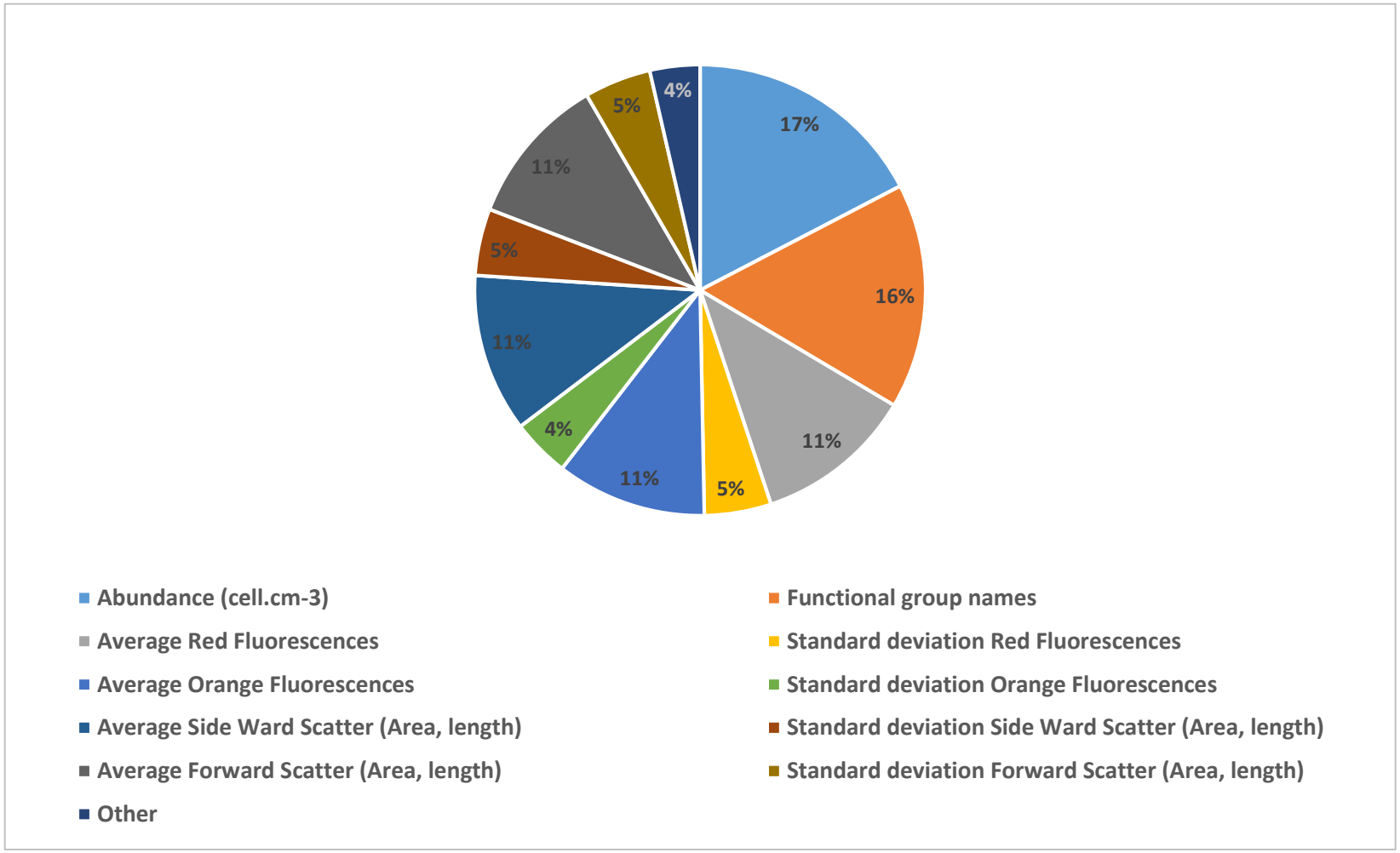


Q: What is the unit used for scatters and fluorescences ?



PART III: Sample Metadata

Q: Which parameters do you export after your clustering?



First FCM ingestion to SDN

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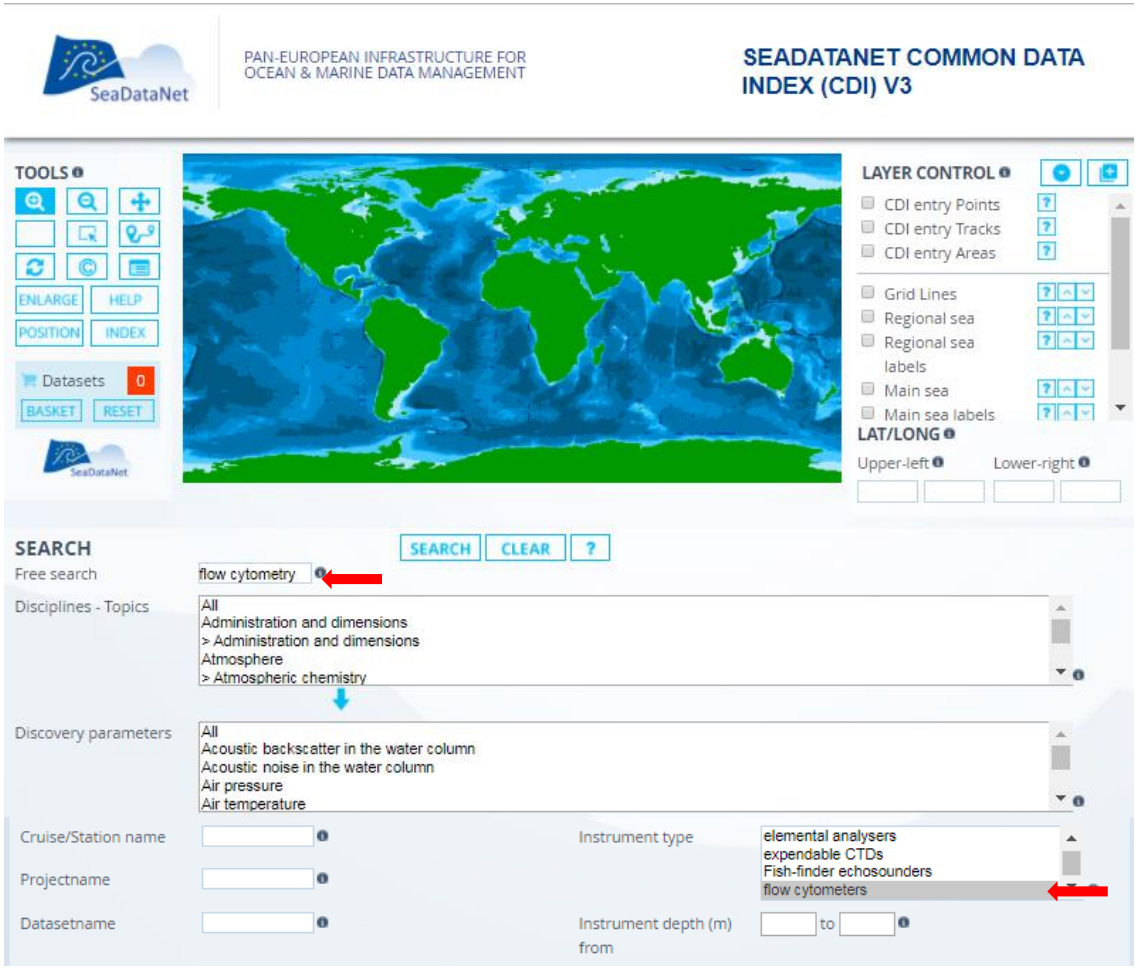
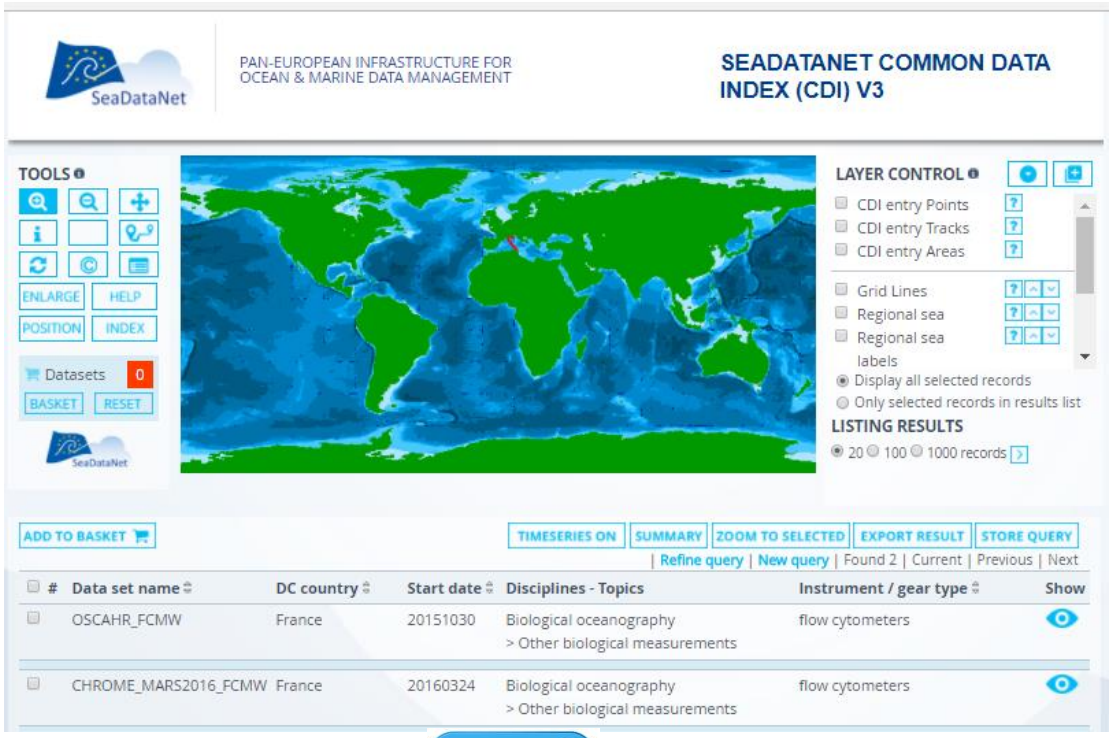


First FCM ingestion to SDN

http://seadatanet.maris2.nl/v_cdi_v3/search.asp



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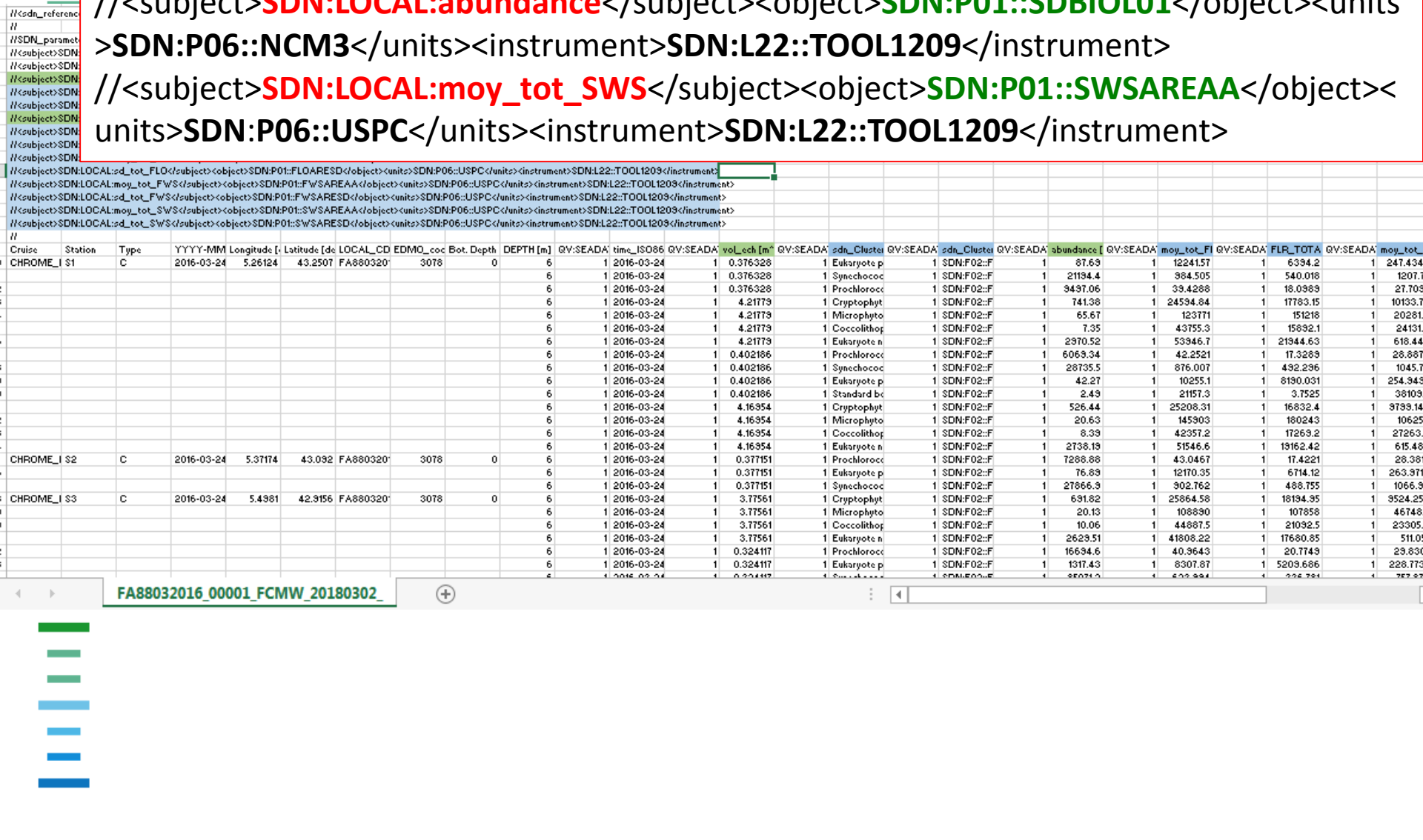

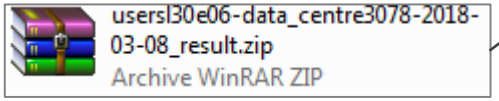
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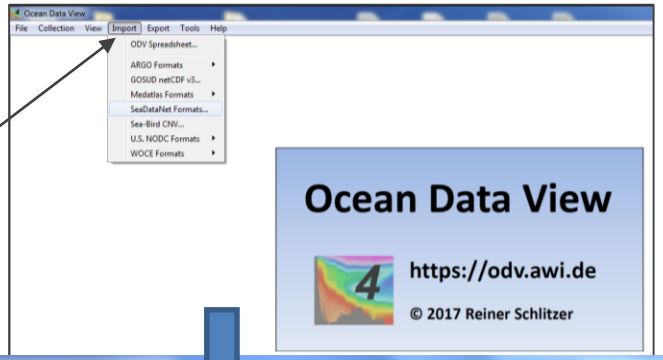
Ocean Data View
File Collection View Import Export Tools Help
ODV Spreadsheet...
ARGO Formats
GOSUD netCDF v3...
Medatlas Formats
SeaDataNet Formats...
Sea-Bird CNV...
U.S. NODC Formats
WOCE Formats

Ocean Data View
https://odv.awi.de
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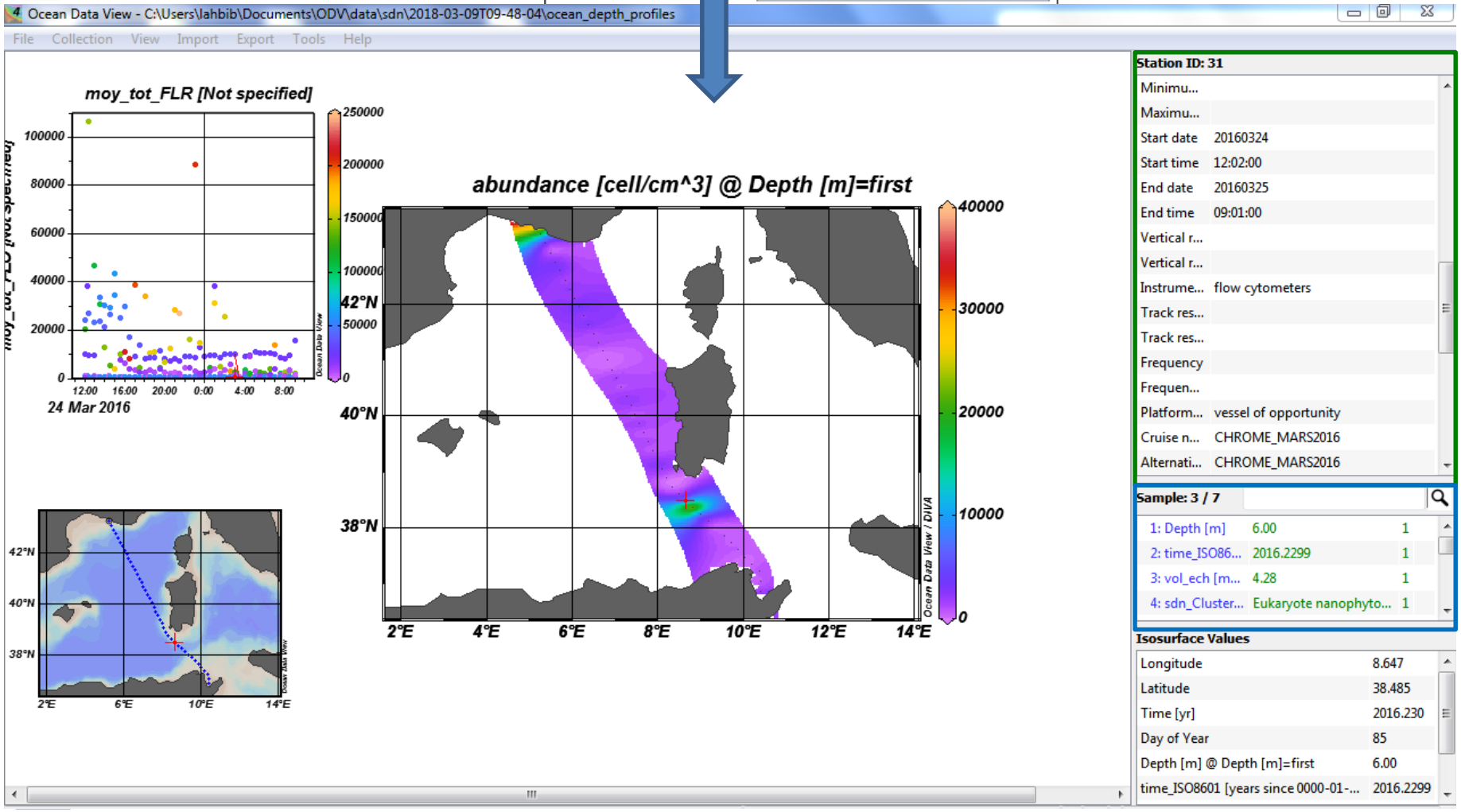
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File Collection View Import Export Tools Help
moy_tot_FLR [Not specified]
24 Mar 2016
abundance [cell/cm^3] @ Depth [m]=first
42°N
40°N
38°N
2°E
4°E
6°E
8°E
10°E
12°E
14°E
Ocean Data View / DIVA
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End time 09:01:00
Vertical r...
Vertical r...
Instrume... flow cytometers
Track res...
Track res...
Frequency
Frequen...
Platform... vessel of opportunity
Cruise n... CHROME_MARS2016
Alternati... CHROME_MARS2016
Sample: 3 / 7
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3: vol_ech [m... 4.28 1
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Latitude 38.485
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Day of Year 85
Depth [m] @ Depth [m]=first 6.00
time_ISO8601 [years since 0000-01-... 2016.2299

Conclusion

- Whatever the instrument used ➔Common Vocabulary (CV)
- We created 44 FCM CV ➔ European portals
- Decide on a group of experts interested in contributing to the vocabulary work and decide on a co-ordinator
- Update is possible/The BODC Vocabulary Editor webpage: https://www.bodc.ac.uk/resources/vocabularies/vocabulary_editor/
- BODC is setting up some repositories on GitHub for each individual collection and F02 will have its own too. So this could be used to share and discuss issues more widely.
- Valuable information are gathered from the questionnaire
- Set up a Quality control protocol for FCM data (70% no flag)

"Needs for standardisation ,
Needs for automated clustering "

Thank you for your attention

This project has received funding from the European Union's Horizon 2020 research and innovation programme under grant agreement No 654410.